

Quickly Synthesized Mercury Nanoparticles from Bacterial Culture Supernatant by Microwave Irradiation

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Abstract: The area of nanotechnology places a premium on the creation of dependable methods for the production of materials on the nanoscale. It has been claimed that microorganisms can synthesize silver nanoparticles, however this process is painfully slow. This paper presents a new method for synthesizing metallic nanostructures of noble metals like silver (Ag) using a combination of *Bacillus subtilis* culture supernatant and microwave (MW) irradiation in water. The process is quick, easy, and "green" because it does not involve a surfactant or soft template. Researchers discovered that silver nanoparticles might be produced by exposing *Bacillus subtilis* culture supernatants to microwave irradiation and silver ions. The size range of the silver nanoparticles was 5-60 nm. In order to study the nanoparticles, researchers used TEM and UV-Visible Spectroscopy. Rapid, chemical-free nanoparticle production is possible with this approach, and the resulting particles retain their stability for months. We conclude that bio-reduction is a viable alternative to electrochemical approaches for producing nanoparticles.

Keywords: Silver nanoparticles, *Bacillus subtilis*, Culture supernatant, Microwave irradiation, Bio-reduction

Introduction

Nanotechnology provides the ability to engineer the properties of materials by controlling their size, and this has driven research toward a multitude of potential uses for nanomaterials. Metallic nanoparticles exhibit unusual optical, thermal, chemical, and physical properties. The reduction of materials' dimension has pronounced effects on the

physical properties that may be significantly different from the corresponding bulk material. Some of the physical properties exhibited by nanomaterials are due to (i) large surface atom, (ii) large surface energy, (iii) spatial confinement, and (iv) reduced imperfections.

One key aspect of nanotechnology concerns the development of rapid and

reliable experimental protocols for the synthesis of nanomaterials over a range of chemical compositions, sizes, high monodispersity and large scale production. A variety of techniques have been developed to synthesize metal nanoparticles, including chemical reduction using a number of chemical reductants including NaBH_4 , N_2H_4 , NH_2OH , ethanol, ethylene glycol and *N,N*-dimethylformamide (DMF)¹⁻⁵, aerosol technique⁶, electrochemical or sonochemical deposition^{7, 8}, photochemical reduction⁹, and laser irradiation technique¹⁰. A lot of interest has been created by the term "green nanotechnology". In a broad sense, this term includes a wide range of possible applications, from nanotechnology-enabled, environmentally friendly manufacturing processes that reduce waste products (ultimately leading to

atomically precise molecular manufacturing with zero waste); the use of nanomaterials as catalysts for greater efficiency in current manufacturing processes by minimizing or eliminating the use of toxic materials (green chemistry principles); the use of nanomaterials and nanodevices to reduce pollution (*e.g.* water and air filters); and the use of nanomaterials for more efficient alternative energy production (*e.g.* solar and fuel cells). With the flourishing demand of “green” nanoparticle synthesis processes¹¹, the field of nanoparticle synthesis has recently developed new routes. These include employing microorganisms, such as: *Pseudomonas stutzeri*¹² *Verticillium sp*¹³, *Fusarium oxysporum*^{14,15}, *Thermomonospora sp*¹⁶ and, also, alfalfa and geranium plants^{17,18}.

Biosynthetic methods employing either biological microorganisms or plant extracts have emerged as a simple and viable alternative to chemical synthetic procedures and physical methods¹⁴⁻²¹. Although it is known that microorganisms such as bacteria, yeast and now fungi play an important role in remediation of toxic metals through reduction of the metal ions, this was considered interesting as nanofactories very recently. The presence of hydrogenase in the *F. oxysporum* broth was demonstrated. Some bacteria reduce Fe^{3+} oxides by producing and secreting small, diffusible redox compounds that can serve as electron shuttle between the microbe and the insoluble iron substrate²². This extracellular enzyme shows an excellent redox properties and it can act as an electron shuttle in the metal reduction. It was evident that electron shuttles or other reducing agents (*e.g.* hydroquinones) released by microorganisms are capable of reducing ions to nanoparticles²³. However, biological nanoparticles synthesis generates nanoparticles at a much slower

rate. In the studies synthesis of silver nanoparticles using the cell mass of bacteria²⁴⁻²⁵ and fungi¹³ or their leached cell components¹⁴, the time required completing the reaction ranged from 24 to 120 h; this lengthy reaction is the one major drawback of the biological synthesis. Biological synthesis would have greater commercial viability if the nanoparticles could be synthesis more rapidly and in the large production scale. Despite stability and a green method of production, particles are not monodispersible and the rate at which they form is not comparable to chemical synthesis methods.

Thermal factors have been demonstrated to affect the size and uniformity of nanoparticles²⁶. Controlling particle growth rate is possible through knowledge of a material's properties and controlling of the reaction by changes to the time and temperature. The ability of any material to absorb microwave energy is expressed by its dielectric loss factor combined with the dielectric constant. The microwave radiation heats up a material through its dielectric loss, which converts the radiation energy into thermal energy. The main

challenges frequently encountered by the researchers of nanoparticle synthesis are

(i) controlling the particle size/shape and
(ii) achieving the monodispersity. The synthesis of nanoparticles with controlled monodispersity is a recent demand by the material developers for the advancement of nanotechnology. Herein, we describe a novel combinatorial synthesis approach which is rapid, simple and “green” for the synthesis of metallic nanostructures of noble metals such as silver (Ag), by using a combination of culture supernatant of *Bacillus subtilis* and microwave (MW) irradiation in water in absence of a surfactant or soft template.

We have chosen *Bacillus subtilis* because these bacteria are ubiquitous bacteria commonly found in various ecological niches including soil, water and air which do not have a history of pathogenicity from contact in the environment.

Chemicals, culture media and test organisms

The bacteria test strain used was *Bacillus subtilis*, (ATCC 6633) strain was obtained from WARD'S Natural Science USA and all other chemicals and the culture media unless otherwise stated were obtained from Merck, Germany.

Experimental

Bacillus subtilis stock cultures were maintained by subculturing at monthly intervals. Growth medium used was Luria Broth (LB) medium (1% Bacto-tryptone [Difco], 0.5% yeast extract [Difco], 1% NaCl, pH 7.5). 250 mL of Luria broth (LB) was prepared, sterilized, and inoculated with fresh batch of the bacteria, *Bacillus subtilis*. The culture flasks were incubated for 36 h at 37°C with shaking at 150 rpm. After incubation period, the cultures were centrifuged at 5,000 rpm for 10 minutes and the supernatant were collected. These supernatant were used as the starting material for synthesis of nanoparticles.

Extracellular biosynthesis of Ag nanoparticles using culture supernatant of bacillussubtilis

In a typical synthesis of silver nanoparticle extracellularly, 50 mL aqueous solution of 1 mM silver nitrate (AgNO_3) was treated with 50 mL of *Bacillus subtilis* supernatant solution in a 250 mL Erlenmeyer flask (pH adjusted to 8.5). The whole mixture was put into a shaker at 40°C (200 rpm) for 5 days and maintained in the dark. Control experiments were conducted with

uninoculated media, to check for the role of bacteria in the synthesis of nanoparticles. The reduction of Ag^+ ions was monitored by sampling an aliquot (2 mL) of the solution at intervals of 24 h. and measuring the UV-Vis spectra of the solution. Absorption measurements were carried out on a Genesys 10 UV-VIS spectrophotometer. The spectra were recorded at room temperature using a one-centimetre quartz cuvette.

The transmission electron microscopy (TEM) analysis of extracellular synthesized silver nanoparticles were prepared by drop-coating biosynthesized silver nanoparticles solution on carbon-coated copper TEM grids (40 μm \times 40 μm mesh size). Samples were dried and kept under vacuum in desiccators before loading them onto a specimen holder. TEM measurements were performed on a JEOL model 1200EX electron microscope operated at an accelerating voltage at 120 kV.

Combinatorial synthesis of nanoparticles using bacterial supernatant and microwave irradiation

As mentioned above, 50 mL aqueous solution of 1 mM silver nitrate (AgNO_3) was treated with 50 mL of *Bacillus subtilis* supernatant solution in a 250 mL Erlenmeyer flask (pH adjusted to 9.0). The whole mixture was put in a domestic microwave oven (National Model

N N-GD 576M). The sample was subjected to several short burst of microwave irradiation at frequency of 2.45 GHz, at power output of about 100W in a cyclic mode (on 15 s, off 15 s) to prevent overheating as well as aggregation of metals. The irradiation process was conducted for a minimum of 5 up to maximum of 15 cycles. The reduction of Ag^+ ions was monitored by sampling an aliquot (2 mL) of the solution after 5, 7, 9, 12 and 15 cycles

and measuring the UV-Vis spectra of the solution. Absorption measurements were carried out on a Genesys 10 UV-VIS spectrophotometer. The spectra were recorded at room temperature using a one-centimetre quartz cuvette. Samples for transmission electron microscopy (TEM) were prepared as described previously.

Nitrate reductase assay

Nitrate reductase converts nitrate to nitrite. The enzyme activity was measured using the method described by Harley³¹ where the activity was measured by putting in the substrate for the enzyme (nitrate) and then measuring the amount of nitrite after 1 h. The net increase in nitrite at 1 h is the amount of nitrate reductase activity. The reagents were: assay medium: 30 mM KNO₃ and 5% propanol in 0.1 M phosphate buffer pH 7.5; nitrite solution: 25 μ M NaNO₂ (Nitrite) solution; nitrite assay reagents: sulfanilamide solution: 1% (w/v) in 25% (v/v) HCl and *N*-(1-naphthyl) ethylenediamine dihydrochloride solution (NEED): 0.02% (w/v) in distilled water.

Five tubes each containing 10 mL of the *Bacillus subtilis* culture supernatant, taken from the 120 h reaction were prepared. Tubes 'a' and 'b' (duplicates) were placed in a boiling water bath sample to kill the enzyme as soon as 10 mL assay medium was added (*i.e.* at time-zero). After boiling 5 minutes, the tubes were kept at room temperature until the nitrite was measured. 10 mL of assay medium were added to tubes 'c, d, e' (triplicates) and incubate in the dark for 60 minutes before placing the tubes in the boiling water bath for 5 minutes. After the samples had cooled, 5 mL of sulfanilamide solution was added to each. Then 5 mL of NEED solution was added and quickly mixed by inverting the tubes. After 20 minutes the absorbance at 540 nm was measured using a Genesys 10 UV-

VIS spectrophotometer. The amount of nitrite found in tubes 'a' and 'b' were subtracted from that in tubes 'c, d', and 'e' to calculate the amount produced by enzyme activity during 60 min. Nitrite standard curve was prepared by placing known amounts of nitrite (0 to 250 nmol NO₂) followed by 5 mL of sulfanilamide and 5 mL of NEED solution. After mixing and incubation for 20 min at room temperature absorbance was measured at 540 nm. The enzyme activity was calculated based on the increase in nitrite over 60 min for the amount of sample (10 mL) and expressed as nmol nitrite/hour/mL.

Determination of the tryptophan/tyrosine residues

Presence of tryptophan/tyrosine residues in proteins released in the bacterial culture supernatant was analyzed spectrophotometrically by measurement of absorbance in the 250 – 300 nm wavelength regions. The measurements were carried out on a Genesys 10 UV-VIS spectrophotometer. The spectra were recorded at room temperature using a one-centimetre quartz cuvette.

Results and Discussion

Extracellular biosynthesis of silver nanoparticles using culture supernatant of *Bacillus subtilis*

Extracellular production of silver nanoparticles by the culture supernatants of *Bacillus subtilis* with aqueous silver nitrate solution, 1mM was investigated. The picture of test tubes of silver nitrate solution after exposure to culture supernatant of *Bacillus subtilis* is shown in the inset of Figure 1. The appearance brown colour clearly indicates the formation of silver nanoparticles in the reaction mixture¹⁹. The characteristic brown colour of colloidal silver solution is due to the excitation of surface plasmon vibrations in the nanoparticle and provides a convenient spectroscopic signature of

their formation.

Several hydroquinones with excellent redox properties were reported that could be act as electron shuttle in metal reductions^{22, 23}. Thus, it was evident that electron shuttle or others reducing agents released by *Bacillus*

subtilis are capable of reducing silver ions to silver nanoparticles. On the other hands, the reduction of silver ions did not occur in the absence of bacterial cells. This clearly indicates that the reducing agents that are released into the cultures of the aforementioned *Bacillus subtilis* are involved in the reduction process.

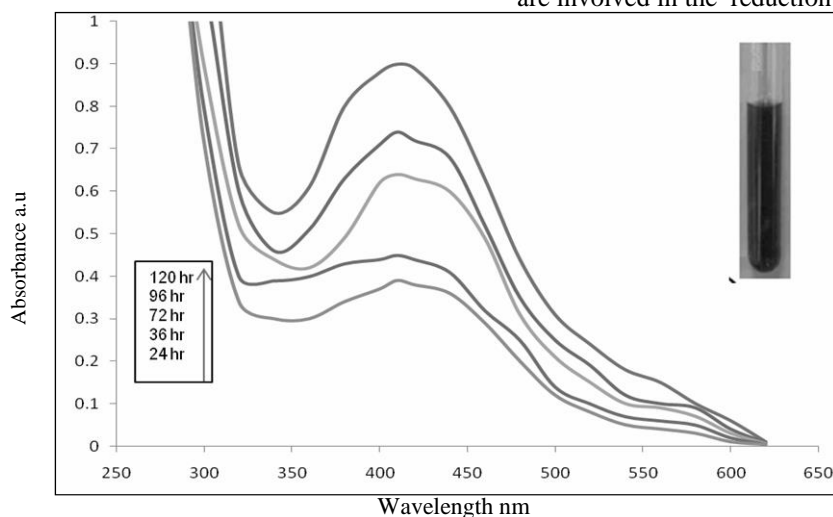


Figure 1. UV-Vis spectra recorded as a function of time of reaction of 1 mM aqueous solution of AgNO_3 with *Bacillus subtilis* culture supernatant. The inset of the figure shows a test tube of the silver nanoparticle solution formed at the end of the reaction.

From the UV-Vis spectra recorded for the aqueous silver nitrate - *B. subtilis* reaction medium as a function of time (Figure 1) it can be observed that the silver surface plasmon band occurs at around 410 nm and steadily increase in intensity as a function of time of reaction. The surface plasmon band in the silver nanoparticles solution remains close to 410 nm throughout the reaction period, suggesting that the particles are dispersed in the aqueous solution with no evidence for aggregation. After completion of the reaction, the silver nanoparticles solution was tested for stability. It was observed that the nanoparticle solution of silver was extremely stable for more than six months with no signs of aggregation even at the end of this period. The particles are thus stabilized in solution by the capping agent that is likely to be

proteins secreted by the biomass.

A representative TEM picture recorded from the silver nanoparticle film deposited on a carbon coated copper TEM grid is shown in Figure 2. This picture shows individual silver particles as well as a number of aggregates. The morphology of the nanoparticles is highly variable, with spherical and occasionally triangular nanoparticles observed in the micrograph. Under observation of such images in an optical microscope, these assemblies were found to be aggregates of silver nanoparticles in the size range 5–50 nm. The nanoparticles were not in direct contact even within the aggregates, indicating stabilization of the nanoparticles by a capping agent. This corroborates with the previous observation by Ahmad *et al*¹⁴ in their study on *F. oxysporum*. As discussed

earlier, the silver nanoparticle solution, synthesized by the reaction of Ag⁺ ions with *Bacillus subtilis* culture supernatant, is exceptionally stable and the stability is likely to be due to capping with proteins

secreted by the bacteria. The separation between the silver nanoparticles seen in the TEM image could be due to capping by proteins and would explain the UV-Vis spectroscopy measurements, which is characteristic of well-dispersed silver nanoparticles.

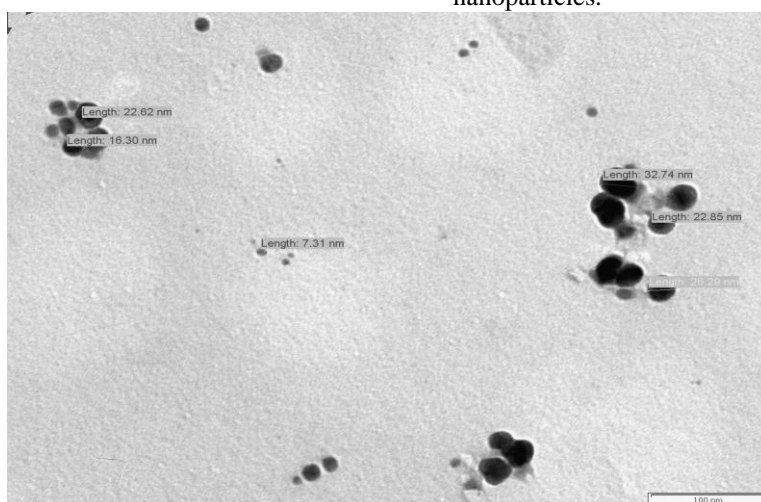


Figure 2. TEM micrograph at 100,000 times magnification recorded from a drop-coated film of an aqueous solution incubated with *Bacillus subtilis* culture supernatant and reacted with 1 mM Ag⁺ ions for 120 h at 40°C. The scale bar corresponds to 100 nm.

Rapid extracellular biosynthesis of silver nanoparticles using culture supernatant of bacillus subtilis and microwave irradiation

The previous method was extended Rapid extracellular production of silver nanoparticles by the culture supernatants of *Bacillus subtilis* with aqueous silver nitrate solution was extended by incorporating microwave irradiation. The advantage of using microwave radiation is that it provides uniform heating around the nanoparticles and can assist the digestive ripening of such particles without aggregation. The microwave radiation heats up a material through its dielectric loss, which converts the radiation energy into thermal energy. From the UV-Vis spectra for microwave irradiated 1 mM AgNO₃ solution in the presence of culture supernatants of *Bacillus subtilis*

(Figure 3) it can be observed that the silver surface plasmon band occurs at around 410 nm and steadily increase in intensity as a function of time of reaction. It can be seen that surface plasmon absorption band of Ag particles was observed just after 5 cycles of irradiation and increased with further irradiation. The absorbance observed at 12th cycle (180 s of irradiation) was roughly similar to that obtained by the non- microwave reaction incubated for

Not only the heating is faster through microwave radiation, but also the temperature distribution of the solution is more uniform. As such, this has led to the fast reaction rate and narrow size distribution of the Ag nanoparticles. This rapid microwave heating also provides

uniform nucleation and growth conditions, leading to homogeneous nanomaterials with smaller sizes. Power dissipation is fairly uniform throughout with “deep” inside-out heating of the polar solvents, which leads to a better crystallinity.

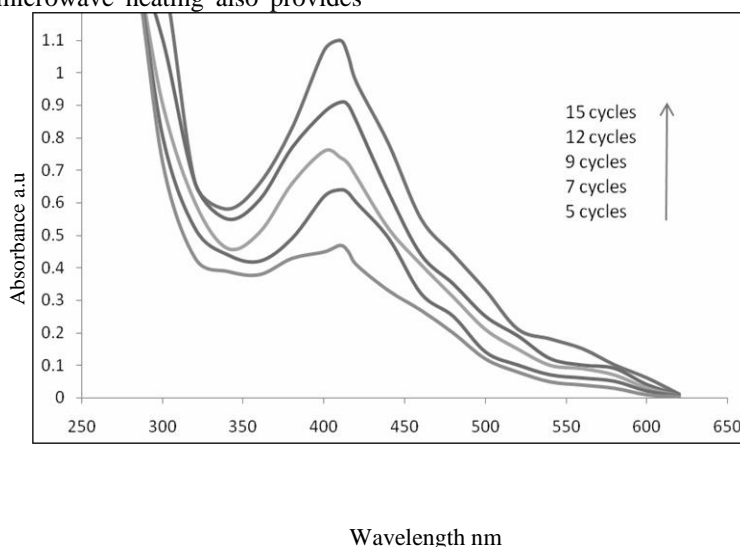


Figure 3. UV-Vis spectra recorded as a function of time of reaction of 1 mM aqueous solution of AgNO_3 with *Bacillus subtilis* culture supernatant irradiated with microwave. The irradiation period was 5 cycles up to 15 (microwave exposure was 15 s per cycle).

It can be seen from Figure 3 that the bands are sharper and more symmetrical, which reflect more uniform size distribution. The major advantage noticed is the time of reaction; the reaction proceeds at a much faster rate when irradiated with microwave.

A representative TEM picture recorded from the silver nanoparticle film deposited on a carbon coated copper TEM grid is shown in Figure 4. This picture shows individual silver particles as well as a number of aggregates. The morphology of the nanoparticles is variable, with majority of them spherical. By gentle heating, using microwave irradiation of an aqueous silver nitrate

solution containing *Bacillus subtilis* culture supernatant, we had obtained relatively mono-disperse silver nanoparticles. Under observation of such images in an optical microscope, these assemblies were found to be aggregates of silver nanoparticles in the size range 5–50 nm. The nanoparticles were not in direct contact even within the aggregates, indicating stabilization of the nanoparticles by a capping agent.

reduction of the Ag^+ ions occurs

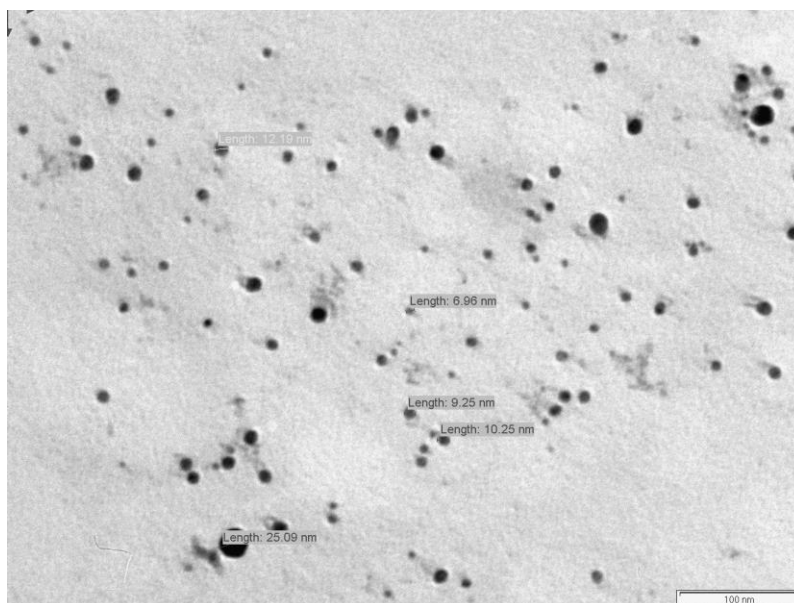


Figure 4. TEM micrograph at 100,000 times magnification recorded from a drop-coated film of 1 mM aqueous solution of AgNO_3 with *Bacillus subtilis* culture supernatant irradiated with microwave. The scale bar corresponds to 100 nm.

The particles are thus stabilized in solution by the capping agent that is likely to be proteins secreted by the biomass. This is an important aspect of synthesis of nanoparticles, since the lack of sufficient stability of many nanoparticles preparation has to some extent impeded the development of the real world applications of nanomaterials.

Application of the biological systems for synthesis of silver nanoparticles has already been reported earlier¹⁶. However, the exact reaction mechanism leading to the formation of silver nanoparticles by all these organisms is yet to be elucidated. Ahmad *et al*¹⁴ have reported that certain NADH dependent reductase was involved in reduction of silver ions in case of *F. oxysporum*. While the above experiments clearly establish that the

extracellularly, it would be important to identify the reducing agents responsible for this. The average protein contains 3.5% phenylalanine, 3.5% tyrosine and 1.1% tryptophan: these three amino acids contribute the majority of the proteins absorbance and fluorescence properties in the 250-300 nm wavelength range, although if the protein contains significant numbers of disulphide bonds these too will contribute to the absorption properties in the wavelength range 250-280 nm. On the other hand the peptide bond and other amino acids contribute to absorbance properties in the 210 – 220 nm regions. The Figure 5 shows the UV-Vis spectrum in low wavelength region recorded from the reaction of 10^{-3} M AgNO_3 with culture supernatant of *Bacillus subtilis* after 12th cycle of microwave irradiation. The presence of the absorption band around 275 nm is attributed to aromatic amino acids of proteins. It is well known that the absorption band at around 275 nm arises due to electronic excitations in tryptophan and tyrosine residues in the proteins. This observation indicates the

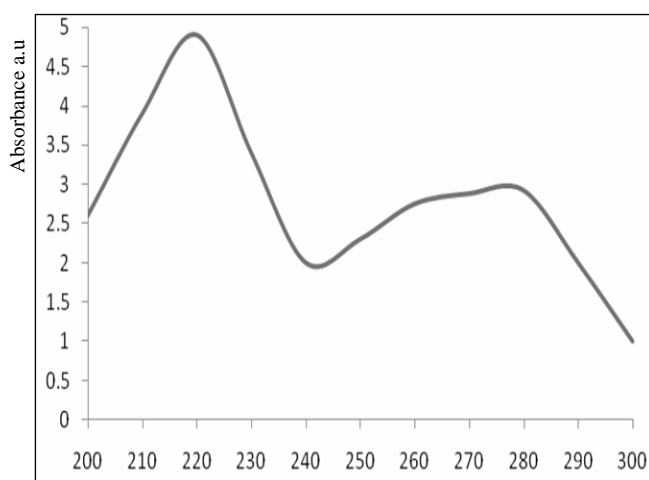
release of proteins into solution by *B. subtilis* and suggests a possible mechanism for the reduction of the metal ions present in the solution. In this study, the UV-Vis spectrum study provides some clues regarding the mechanism of synthesis of

silver nanoparticles. There were two absorbance peaks found in the UV range corresponding to 220 and 275 nm. While the peak at 220 nm may be due to absorption by amide bond, the other peak at 275 nm may be attributed to the tryptophan and tyrosine residues present in the protein.

This indicates secretion of some proteinic components into the medium by the bacterial biomass which might play important role in the reduction of the metal ions in the form of nanoparticles. Consequently the proteins also may bind to the nanoparticles and enhance the stability.

Wavelength nm

Figure 5. UV-Vis spectra in the low wavelength region recorded from the reaction of 10^{-3} M AgNO_3 with culture supernatant of *Bacillus subtilis* after 12th cycle of microwave irradiation.



Previous studies^{14-16, 19} have indicated that NADH- and NADH-dependent enzymes are important factors in the biosynthesis of metal nanoparticles. The reduction seems to be initiated by electron transfer from the NADH by NADH-dependent reductase as electron carrier. Many fungi that exhibit these characteristic properties, in general, are capable of reducing Au(III) or Ag(I)²⁹. Besides these extracellular enzymes, several naphthoquinones³¹ and anthraquinones²³ with excellent redox properties, were reported in *F. oxysporum* that could be act as electron shuttle in metal reductions²². The exact mechanism of the reduction of metal ions is yet to be elucidated for bacteria. In this study nitrate reductase activity was detected in the culture supernatant of *Bacillus subtilis* and was about 152 nmol/ h/ mL culture supernatant of *Bacillus subtilis*. It appears that the reductase together with electron shuttling compounds and other peptides/proteins may be responsible for the reduction of Ag⁺ ions and the subsequent formation of silver nanoparticles.

In conclusion, the *Bacillus subtilis* has shown potential for extracellular synthesis of fairly monodispersed, silver nanoparticles in the range of 5–50 nm. The kinetics of silver nanoparticles synthesis using the cell filtrates in combination with microwave irradiation indicates that the rapid synthesis of nanoparticles would be suitable for developing a “green nanotechnology” biosynthesis process for mass scale production. The extracellular synthesis offers a great advantage over an intracellular process of synthesis from the application point of view. Since the nanoparticles formed inside the biomass would have required additional step of processing for release of the nanoparticles from the biomass by ultrasound treatment or by reaction with

suitable detergents. The extracellular synthesis of nanoparticle makes it possible to harness and immobilize/deposit onto desired solid support for the use of different practical purposes. In future, it would be important to understand the biochemical and molecular mechanism of the synthesis of the nanoparticles by the cell filtrate in order to achieve better control over size and polydispersity of the nanoparticles

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